

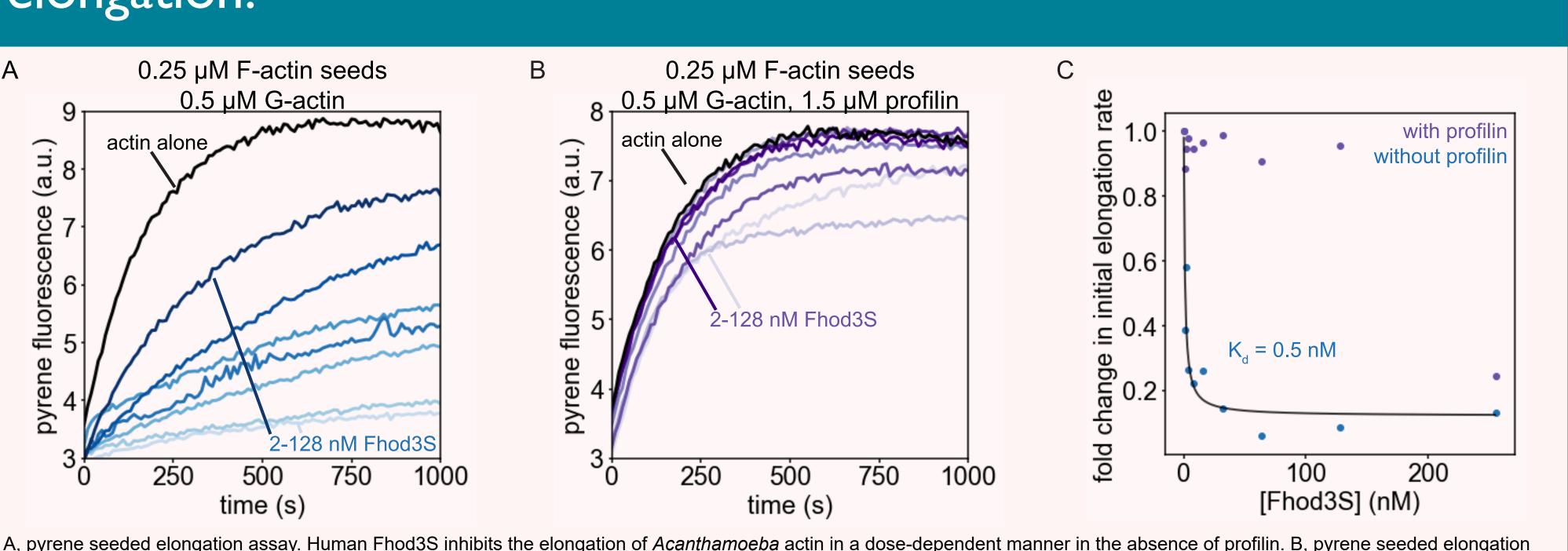
assemble actin in cardiomyocytes?

Actin assembly by Fhod3 in the cardiac sarcomere

Aanand A. Patel¹, Alex M. Grunfeld², Haruko Nakano³, Andrea Chaney², Atsushi Nakano^{3,4,5}, and Margot E. Quinlan^{2,4} ¹Molecular Biology Interdepartmental Doctoral Program, ²Department of Chemistry and Biochemistry, ³Department of Molecular, Cell, and Developmental Biology, ⁴Molecular Biology Institute, ⁵Eli and Edythe Broad Center of Regenerative Medicine and Stem Cell Research; University of California, Los Angeles

A, pyrene actin assembly assays. Human Fhod3 (both the muscle-specific Fhod3L isoform and the non-muscle Fhod3S isoform) nucleate actin from rabbit skeletal actin. B, pyrene actin assembly assays. Fhod3 splice isoforms nucleate human platelet actin (80% β-actin, 20% γ-actin). C, quantification of actin assembly rates at time to half-polymerization from A and B. Fhod3 polymerizes Acanthamoeba actin faster than it does mammalian actins.

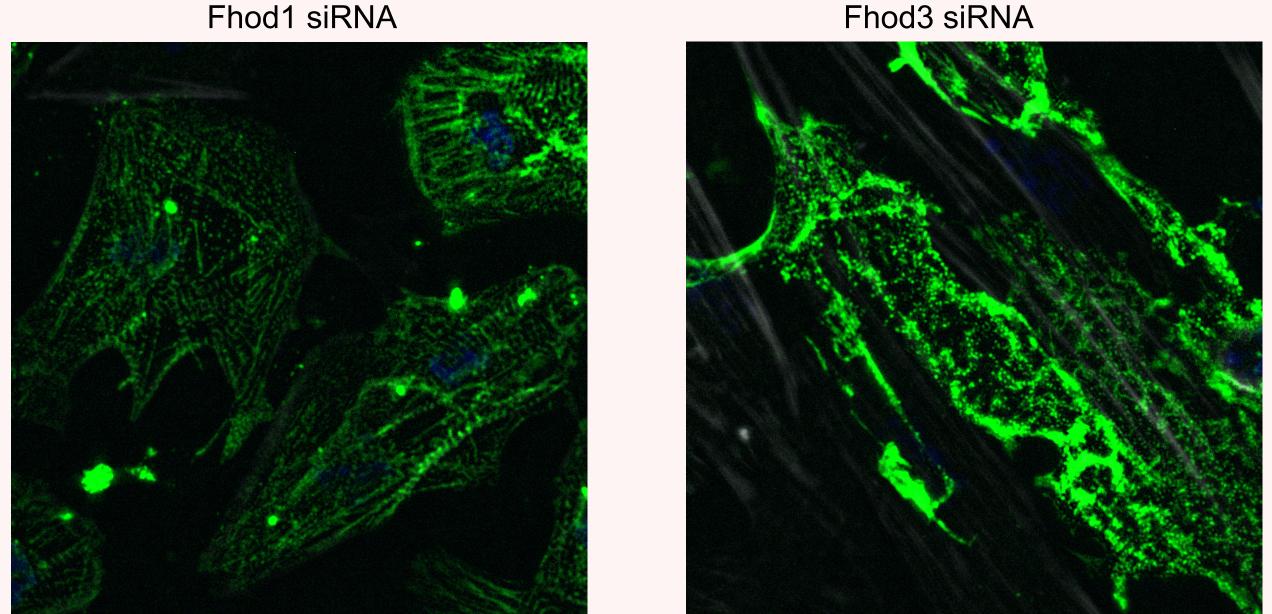
3. Human Fhod3 binds barbed ends but does not accelerate elongation.



A, pyrene seeded elongation assay. Human Fhod3S inhibits the elongation of Acanthamoeba actin in a dose-dependent manner in the absence of profilin. B, pyrene seeded elongation assay with S. pombe profilin. Human Fhod3S permits, but does not accelerate, the elongation of Acanthamoeba actin in the presence of profilin. C, quantification of rates from A and B. Fhod3S binds barbed ends of *Acanthamoeba* actin with an affinity of ~0.5 nM.

4. Fhod3 is required for sarcomere assembly in hESC-derived cardiomyocytes.

scramble siRNA



Human embryonic stem cell-derived cardiomyocytes were transfected with scramble, Fhod1, or Fhod3 siRNA, then fixed 21 days post-differentiation and stained for α-actinin. Depletion of Fhod3, but not Fhod1, led to a complete loss of sarcomeres, consistent with previous work in other model systems.

Future Directions

1. Systematically measure the nucleation and elongation rates of Fhod1, Fhod3L, and Fhod3S with each actin and profilin isoform.

2. Map the sites in Fhod1 and actin that confer isoform specificity.

3. Generate mutants and chimeras that alter nucleation, elongation rate, and isoform-specificity of Fhod1 and Fhod3.

4. Test ability of mutants to assemble actin in human cardiomyocytes.

Fhod1 siRNA

Acknowledgements

We thank members of the Quinlan and Reisler labs for their feedback and UCLA Graduate Division for travel funding. This work was supported by the following NIH grants: NRSA F30 HL137263, MSTP T32 GM008042, and R01 GM096133.

References

Al Haj et al., 2014. Eur. J. Cell Biol. Bennett et al., 2006. Mol. Biol. Cell Benz et al., 2013. Cell Commun. Signal. Craig and Pardo, 1983. Cell Motil. Dwyer et al., 2014. Anat. Cell (Hoboken) Fenix et al., 2018. *bioRxiv* Goode and Eck, 2007. Annu. Rev. Biochem. Hatano et al., 2018. J. Cell Sci. Iskratsch et al., 2010. J. Cell Biol. Patel et al., 2018. J. Biol. Chem. Schönichen et al., 2013. J. Cell Sci. Taniguchi et al., 2009. J. Biol. Chem.